

IMCB Flow Core Facility		Prepared by:	IMCB Flow Core Facility				
Cell Analysis/Sorting Form		Email:	imcb_flowcytometry@a-star.edu.sg				
		Tel: Effective from:	65-6586 9830 14 Feb 2020				
Customer Information:			14100 2020				
PI Name:	PI	Email:					
Department & Institute:	I						
Billing Address:							
Requestor Name:		Requestor's					
Requestor Email:		Phone:					
Fund/Grant Code (A-STAR):		WBS Accoun (A-STAR):	t				
Date/Time service required: (dd/mm/yyyy)	(/ /)	2	🛆 Afternoon (🛛 / 🕦				
Operating hours: week days 10am-1pm (morning) and 2pm-6p	om (afternoon)						
Sample Declaration (Requestor to provide infor	mation):						
Are any of the samples:							
1. Of human origin?	Please indicate species, source and clinical history of specimen.						
Yes 🛆 No	Species:						
	Source:						
2. Of non-human primate origin?							
Yes 🛆 No	Clinical History:						
a. If yes, are the animals negative for Herpesvirus B or SIV (Simian Immunodeficiency Virus) infection? Yes 🛆 No	If involving human samples, IRB Status : Approved/Exempted/N.A If Approved, IRB Approval number :						
 Obtained from human subjects undergoing febrile illness of unknown origin? 	4. Infected with H Immunodeficiency (Hepatitis C Virus)	HV (Human Virus), HCV or any known	5. Characterized by active viral replication (eg. <i>In vitro</i> infected cell lines)?				
	Biosafety Level 3 pathogen?		Yes 🛆 No				
Yes 🛆 No	Yes Δ M	No					
6. Transduced with plasmid or viral vectors? *If Q5 or Q6 is ves. please specify vector and/or infecting virus.							
Yes 🛆 No							
	L						
355nm (UV): 🛆 BUV737 🛆 BUV496 🛆 BUV395							
405nm (Violet): 🛆 BV786	6 \triangle BV711 \triangle Qdot655/BV650 \triangle BV605 \triangle BV480 \triangle BV421						
Colors: 488nm (Blue): Δ FITC/Alexa488/GFP Δ YFP Δ PerCP-Cy5.5/7AAD							
561nm (Yellow-green): 🛆	w-green): Δ PE Δ RFP/mCherry/PE-CF594 Δ PE-Cy5 Δ PE-Cy7						
635nm (Red): 🛆 APC/Alex	d): \triangle APC/Alexa647 \triangle APC-Cy7						

	Service(s) Provided (to be filled by FACS operator):							
	Cell analysis / sorting prices (excluding 7% GST) (Usual rate: \$130 per hour)							
	Date	Start Time	End Time	Total No. of Hours	Unit Price (\$)	Total Cost (\$)		
Analysis (Aurora/LSRII)					50.00			
Analysis (Symphony)					80.00			
Sorting (Aria)					115.00			
					Grand Total (excl			
					GST)			
	Confirmation of Order:							
	I hereby declare that the information provided above is correct and that I have read and understood the terms and conditions for using the cell-sorting facility. FACS Operator / Date FACS Operator / Date							
	Acknowledgement of	Services:						
	Service Completion Date (dd/mm/yyyyy):							
	I hereby acknowledged that the service has been completed.							
	FACS Oper	ator / Date		Signature o	f Requestor / Date			

For Internal Reference Only				
Billing Information:				
Invoice Number/Date:	Payment Date:			
Verified By:				

Terms and conditions:

- 1. Contact IMCB Flow Core at <u>imcb_flowcytometry@imcb.a-star.edu.sg</u> to plan <u>a</u> sort every time you want to do a new type of experiment to discuss the requirements and set up of the machine.
- 2. Operation of the machine is strictly performed only by the core facility operators for sorting.
- 3. Charges for the use of this facility is as stipulated above, excluding GST.
- Sorts need to be scheduled at least 5 days before. A full cancellation fee applies for cancellations made less than
 24 hrs prior to usage. Users have to send a cancellation email to <u>imcb_flowcytometry@imcb.a-star.edu.sg</u> at least
 24 hours prior to the time of the booking (preferably earlier so we can reschedule).
- 5. Please notify us as soon as possible if there are changes to the sort details provided in the form.
- 6. Requestors are responsible for providing:
 - The necessary controls to ensure the proper gating and identification of the desired cell populations.
 - Collection tubes containing the appropriate media for the sorted cells.

Sample preparation guidelines:

- 7. Samples have to be filtered through a 40 μ m cell strainer to prevent clogging of the nozzle.
- 8. For standard sorts, a concentration of 10 million cells/ml is recommended (3-20 million cells/ml is the tolerable range for sorting).
- 9. Final cell yield is dependent upon many factors: starting percent of target population, total number of live and dead cells in the sample, cells stickiness, etc.
- 10. For improved cell yield, collection tubes can be pre-blocked overnight with media containing 10% FCS. The use of polypropylene instead of polystyrene tubes for collection can help to reduce cell stickiness.
- 11. Use of antibiotics in the collection media is recommended to prevent contamination but requestors should determine whether it is appropriate based on its effect on the sorted cell population.
- 12. Important information to note:
 - Size of your cells: to select the proper nozzle size
 - Cell Adhesiveness: to select resuspension buffer to minimize clogging
 - Total cell number: to achieve optimal sorting concentration (minimal clumping, maximum speed)
 - Approximate percentage of your target population: to determine sorting time and maximum possible yield

Biological hazards:

- 13. All potentially biohazardous sorts and protocols will have to be pre-approved by PI in charge.
- 14. Due to biosafety reasons we will not sort:
 - Samples containing lentiviral vectors
 - Radioactive or radioactive labelled samples
 - Human samples
 - i.Undergoing a febrile illness of unknown origin
 - ii.Infected with HIV, HCV or any BSL3 pathogen
 - iii.Characterized by active viral replication (eg. In vitro infected cells)
 - Mammalian / murine samples
 - i.Undergoing infections of unknown origin
 - ii.If infected by human pathogens, the same rules described above will apply
 - iii.Non-human primate cells infected with Herpesvirus B or Simian Immunodeficiency Virus