

Clinical update

Cardiac endothelium-myocyte interaction: clinical opportunities for new heart failure therapies regardless of ejection fraction

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Heart failure (HF) is an important global health problem with great socioeconomic burden. Outcomes remain sub-optimal. Endotheliumcardiomyocyte interactions play essential roles in cardiovascular homeostasis, and deranged endothelium-related signalling pathways have been implicated in the pathophysiology of HF. In particular, disturbances in nitric oxide (NO)-mediated pathway and neuregulin-mediated pathway have been shown to contribute to the development of HF. These signalling pathways hold the potential as pathophysiological targets for new HF therapies, and may aid in patient selection for future HF trials.

Keywords

Heart failure • Endothelium-cardiomyocyte interactions • Nitric-oxide mediated pathway • Neuregulin-mediated pathway • Therapy

Heart failure is a global health problem

Heart failure (HF) is a major global public health problem, with 26 million sufferers worldwide. Heart failure is the top cause of hospitalizations in those \geq 65 years old, with rates of hospitalization on the rise globally. There is a high burden of re-admission¹ and mortality.² Multiple studies have demonstrated substantial improvement in HF survival since the late 1990s,^{3,4} but outcomes remain dismal particularly for the half of the HF population with HF with preserved ejection fraction (HFPEF),⁵ for which there is still no proven therapy. Given the increasing global burden of HF and the projection that HFPEF will become the dominant form of HF in ageing societies, there is an urgent need to explore newer therapies applicable to both HF with reduced ejection fraction (HFREF) and HFPEF.

The cardiac endothelium in cardiovascular homeostasis

In the pathophysiology of HF, a pivotal role has been attributed to generalized endothelial dysfunction, in particular in the cardiac, pulmonary, renal, and muscular endothelial vasculature (*Figure 1*). Importantly, dysregulation of the communication between cardiac endothelial cells and cardiomyocytes has been implicated in the development of cardiac structural and functional abnormalities, and the restoration of endothelium-related signalling pathways is an emerging new approach to mitigate the progression of HF. Importantly, the cardiac endothelium which directly communicates with subjacent cardiomyocytes in the cardiac capillaries and at the endocardium, should be distinguished from the vascular endothelium in the coronary arterial system. Similarly, as in the rest of the systemic circulation, the latter coronary vascular endothelial cells merely

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Figure 1 Schematic presentation of the endothelial system in the organism, and its cell-to-cell interactions in different compartments of the circulation. The endothelium is not commonly thought of as a system and rather studied according to the interests of a specific clinical organ-oriented specialty. In cardiology, most attention has been directed to the (coronary) vascular endothelium in vasomotor control, and to the pathogenesis and management of coronary atherosclerosis and myocardial ischaemia. Yet the arterial endothelium comprises only a minute fraction of the endothelial surface. On the other hand, capillary endothelial cells which directly overlie organ-specific tissue cells, account for the largest number of endothelial cells in the circulation. The role of capillary endothelial cells has long been underestimated, except for the interactions with astrocytes and neurons in the brain. In particular, the interactions between capillary endothelial cells with adjacent organ cells, such as hepatocytes and Kupffer cells in the liver, tubular and glomerular epithelial cells in the kidney, alveolar epithelial cells in the lungs, or myofibre cells in skeletal muscle are likely to be important but have scarcely been studied. In the heart, endothelial cells and adjacent cardiomyocytes communicate in an obligatory fashion, inherent to (i) the anatomical proximity of endothelial cells and cardiomyocytes, both in the endocardium and at the level of the microcapillaries, (ii) the adjustable release of various cardioactive diffusible factors by endothelial cells is a common pathophysiological step during many diseases and leads to ventricular dysfunction, pulmonary and systemic hypertension, kidney failure, and exercise intolerance.

conduct and mediate perfusion of the heart through interaction with subjacent smooth muscle.

The heart is a highly organized pluricellular organ consisting of different, equipotent cell types including endothelial cells, fibroblasts, and cardiomyocytes. Communication between these cell types in a network structure is crucial for cardiac development, autoregulation, and adaptation.^{6,7} The concept that cell–cell communication in the heart is important for cardiac performance was formulated almost 30 years ago by Brutsaert *et al.*,⁸ from experimental observations that selectively damaging the endocardial or microvascular endothelium of isolated papillary muscles influenced the mechanical performance of the underlying cardiac muscle.^{9,10} To date, a growing number of endothelial-derived cardio-active factors have been identified (*Figure 2*), including nitric oxide (NO), endothelin-1, neuregulin-1 (NRG-1), angiotensin II, angiopoietins, prostaglandins, connective tissue growth factor, fibroblast growth factor, vascular endothelial growth factor, Dickkopf-3, apelin, and endothelial miRNAs (*Table 1*). Importantly, half-life and/or duration of signalling activity of these substances appear to vary, reflecting a specific modulatory time-window for each of these factors. This window may be long and sustained (e.g. for most growth factors) or can be very brief, confining modulation to a brief interval in the cardiac cycle. In the latter case, the moment of release must be tightly coordinated within the cardiac cycle, as is the case for NO.¹¹

Recent experimental observations and pharmacological developments have identified pharmacological tools and clinical opportunities to treat HF based on the concept of endothelium– cardiomyocyte interactions. To date, two endothelial pathways, the NO-mediated pathway and the neuregulin-mediated pathway, have progressed to the stage of clinical trials in human HF patients. We aim to review these endothelial pathways and discuss their potential clinical applications in this paper.



Figure 2 Cardiac endothelium–cardiomyocyte communication. Cardiac endothelial cells (in endocardium and myocardial micro-capillaries) secrete many factors in an adjustable fashion, including proteins and diffusible substances like nitric oxide (NO). The cardiac activity of some of these factors like nitric oxide, prostacyclin (PGI2), endothelin-1 (ET-1), and neuregulin-1 (NRG-1) has been recognized. For many other factors, like Dickkopf-3 (DKK3), periostin, TSP-1 (thrombospondin-1), follistatin (FST), apelin, connective tissue growth factor (CTGF) effects on the heart have only emerged recently. The synthesis, secretion, and activities of these factors are closely linked, interrelated, and interactive. Most factors modulate the actions of factors on the same target cell, actions which may be mutually additive, synergistic, or inhibitory. They may even result in novel effects, not seen with either agent alone, also influenced by the milieu in which the interactions occur. With this background, targeting cardiac endothelial function with a (pharmacological) intervention which abruptly imposes dominancy of one of these intertwined pathways may be risky. A gentle, gradual, balanced, and sustained rehabilitation of endothelial function appears a preferable goal.

Table I	Overview of selected endothelium-derived factors with modulatory effects on cardiomyocyte function	

Factor	Receptor/Pathway	Contractility/Relaxation	Hypertrophy/Heart failure	References
Nitric oxide	sGC	+ and – Inotropic effects Increased relaxation	Attenuates cardiac remodelling	15,27
Endothelin-1	ET _A ET _B	Mainly + inotropic Increased relaxation	Increases hypertrophy and fibrosis	127
Prostacyclin	PG-I ₂ receptor	+ and - Inotropic effects	Anti-hypertrophic	7,128,129
Neuregulin-1	Erbb4	Reduces contractility	Attenuates HF progression	130,131
Periostin	lpha-V/ eta -3 and $lpha$ -V/ eta -5 integrins	?	Increases fibrosis and myocyte proliferation	132,133
Thrombospondin-1, -2, -4	Cell adhesion receptors	Role in stretch mediated contractility augmentation (TSP-4)	Anti-hypertrophic Anti-fibrotic	134–137
Follistatin and follistatin-like factor (Fstl1)	Antagonists of TGF-β superfamily cytokines	?	Anti-apoptotic (Fstl1) Anti-hypertrophic (Fstl1 and 3)	138–141
Connective tissue growth factor (CTGF)	Integrin receptors	?	Pro-fibrotic Pro-hypertrophic	142,143
Dickkopf-3 (DKK-3)	Wnt signalling	?	Anti-hypertrophic	144
Apelin	Apelin receptor	+ Inotropic	Anti-hypertrophic	145–147
miR-146a	16K Prolactin pathway	?	Induces peripartum cardiomyopathy	148

The nitric oxide-soluble guanyl cyclase-cyclic guanosine monophosphate pathway

Nitric oxide plays an important role in cardiovascular homeostasis

Initially thought to exert its effects primarily on the vasculature, including on the coronary vasculature, it is increasingly recognized that NO plays an obligatory role in the regulation of myocardial function via its autocrine and paracrine effects. NO is synthesized from the amino acid L-arginine, through a process catalysed by NO synthase (NOS). Expressed in both constitutive [endothelial (eNOS), neuronal (nNOS)] and inducible (iNOS) forms, it can be found in the cardiac endothelium (eNOS, iNOS), myocardium (eNOS, nNOS, iNOS), nerve fibres (nNOS), and inflammatory cells (iNOS).

The fundamental signalling pathway of NO is via stimulation of soluble guanyl cyclase (sGC), leading to the production of the second messenger cyclic guanosine monophosphate (cGMP). Depending on the site of cGMP activation and the site of NO release, the consequent biological actions are different.

Nitric oxide and natriuretic peptides contribute to spatially distinct cyclic guanosine monophosphate pool

Apart from the NO-sGC-cGMP pathway, synthesis of cGMP is also regulated by the natriuretic peptide (NP)-particulate GC (pGC)-cGMP pathway. Initially thought to be contributing to a common cGMP pool, studies have shown that cGMP produced by pGC and sGC is compartmentalized to distinct sub-cellular regions, leading to different responses.^{12,13} NP activation leads to increased cGMP production around the sub-sarcolemmal region, whereas NO signalling results in an increase in the cytosolic pool of cGMP. The spatial boundaries of the distinct cGMP pools are poorly defined, restricted by the activity of cGMP-hydrolysing phosphodiesterases (PDE); PDE-2 limits the sub-sarcolemmal cGMP pool and PDE-5 controls the cytosolic cGMP pool.^{12,14} In addition to compartmentalization, these cGMP pools appear to be functionally distinct. While cGMP produced by pGC and sGC enhances myocardial lusitropy, only cGMP produced by sGC blunts myocardial response to β -adrenergic stimulation.¹³

Nitric oxide and cardiac function

NO modulates cardiac function through its inotropic, lusitropic, and chronotropic effects. It is generally accepted that NO exerts bimodal effects on inotropy.^{15,16} At low and physiological NO concentrations, the NO-sGC-cGMP pathway enhances inotropy via activation of protein kinases A (PKA) and G (PKG) which increases Ca^{2+} concentration.¹⁷ Higher amounts of NO result in negative inotropy;¹⁸ mediated by PKG, the increased cGMP production leads to blockade of sarcolemmal Ca^{2+} channels and reduction in the sensitivity of troponin C to Ca^{2+} . The definitions of 'low' and 'high' concentrations are controversial, given that the amounts of NO produced *in vivo* may not correspond to amounts of bioactive NO delivered. It is therefore not surprising that not all studies demonstrated these bimodal effects on inotropy. At higher concentrations, NO also enhances lusitropy

by accelerating myocardial relaxation and improving myocardial distensibility.^{15,17} Phosphorylation of troponin I reduces cardiac myofilaments' sensitivity to calcium and promotes diastolic cross-bridge detachment.¹⁹ Phosphorylation of titin by PKA and/or PKG increases its compliance, hence improving lusitropy.²⁰ The challenge is identifying the 'sweet spot' where lusitropy is enhanced without excessive compromise on inotropy.

In addition to these direct actions on cardiac function, the general vasodilatatory properties of the NO pathway may also indirectly contribute to cardiac function by altering cardiac pre- and afterload conditions. In particular, an altered afterload of the left ventricle (LV) will, through changes arterial impedance, modulate LV systolic duration and the timing of arterial reflected waves, and thereby optimize the crosstalk between the LV and peripheral circulation without necessarily affecting arterial systemic blood pressure.^{21,22}

The chronotropic effects of NO differ according to the site of action. Animal studies demonstrated positive chronotropic effects via cGMP-dependent stimulation of the hyperpolarization-activated pacemaker current (l_f).²³ Yet, experiments using isolated neonatal and adult ventricular myocytes demonstrated negative chronotropic effects, modulated at the post-synaptic level.²⁴

Through effects on mitochondrial respiration, NO promotes free fatty acids as the preferred myocardial energy substrate and protects against excessive oxygen consumption. In canine models of experimental HF, a drop in myocardial NO production was associated with a shift in myocardial substrate utilization to glucose.²⁵

Role of nitric oxide in heart failure

There is increasing recognition of the central role that NO imbalance plays in contributing to the abnormal cardiac and vascular phenotypes seen in chronic HF, regardless of EF.

HFREF, with post-myocardial infarction as the prototype, is characterized by eccentric remodelling and LV dilatation. In post-infarct models of HFREF, there was a greater degree of adverse remodelling in NOS knock-out mice, compared with wild-type mice; this was mediated via the NO signalling pathway.²⁶

In HFPEF, concentric LV remodelling with increased LV wall thickness is thought to be driven by a pro-inflammatory state, which results in increased oxidative stress through the activation and migration of leucocytes and increased superoxide production. NO is then diverted to peroxynitrite by its interaction with superoxide.²⁷ Low bioavailability of NO and high peroxynitrite levels result in low intracellular cGMP and reduced PKG activity.^{28,29} A deficient NO-sGC-cGMP pathway increases diastolic cytosolic Ca²⁺ and delays myocardial relaxation. Myocardial stiffness also increases with a deficient NO-sGC-cGMP pathway since there is hypophosphorylation of titin via PKG or PKA, with resultant decreased compliance. Accordingly, rodent studies showed that LV diastolic dysfunction and leftward shift of the LV pressure-volume relationship occurred when NO synthesis was inhibited.³⁰ Conversely, infusions of NO donors improved LV diastolic compliance via phosphorylation of titin.³¹ If this trial could be repeated in multicentre studies including a large number of patients, it could be a major breakthrough in the treatment of patient with HFPEF. Beyond the direct actions on the myocardium, NO-sGC-cGMP deficiency may also create a mismatch of the crosstalk between the LV and the systemic arterial impedance. Moreover, it also underlies the vasomotor dysfunction in systemic, pulmonary, renal, muscular, and coronary circulations. Pulmonary vascular endothelial dysfunction with deficient NO-sGC-cGMP signalling leads to pulmonary vasomotor dysfunction, vascular remodelling, and pulmonary hypertension.³² This secondary pulmonary hypertension directly affects right-ventricular (RV) function.³³ NO deficiency also drives renal vasomotor dysfunction, contributing to the development of cardiorenal syndrome (concomitant cardiac and renal dysfunction).³⁴ In the coronary circulation, NO-sGC-CMP deficiency causes inappropriate coronary vasoconstriction, triggers smooth muscle proliferation and vascular remodelling, and reduced myocardial perfusion reserve.³⁵

Propagation of heart failure by worsening nitric oxide deficiency

The neurohormonal activation and systemic inflammation in HF result in endothelial dysfunction, down-regulation of eNOS expression, uncoupling of eNOS, and increased production of reactive oxygen species.³⁶ Chronic inactivity in HF, with altered local endothelial shear stress, further down-regulates eNOS. In addition, the altered redox state causes NO resistance by reducing the levels of NO-sensitive sGC. sGC exists in two different forms, the NO-sensitive reduced sGC and NO-insensitive oxidized sGC. In HF, the balance is shifted in favour of oxidized sGC with reduced NO-sGC-cGMP signalling activity. These eventually culminate in dysfunctional endothelium with attenuated NO activity, which in turn propagates the progression of HF.

Endothelial dysfunction is prevalent in heart failure and portends a worse prognosis

Endothelial dysfunction is highly prevalent in HF, regardless of EF. It was first demonstrated in HFREF patients two decades ago. Since then, there has been a wealth of data showing endothelial dysfunction among HFREF patients in various circulatory beds, and involving large conduit³⁷ and small resistance vessels,³⁸ in addition to the microvasculature.³⁹ Endothelial dysfunction independently predicts adverse cardiovascular outcomes⁴⁰ and correlates with the extent of exercise intolerance.⁴¹ Similarly, endothelial dysfunction was present in 42% of HFPEF patients (compared with 28% of hypertension and none of healthy controls), and correlated with the extent of exercise intolerance⁴² and pulmonary hypertension.⁴³ Akiyama et al.⁴⁴ extended this finding, by further demonstrating that peripheral endothelial dysfunction independently predicted cardiovascular events. The high prevalence and independent prognostic impact suggest that endothelial dysfunction plays an important pathophysiological role in HFPEF.

The neuregulin-erythroblastic leukaemia viral oncogene homolog pathway

Neuregulins and their receptors

NRG-1 is a growth factor belonging to the epidermal growth factor (EGF) family, expressed in the nervous system, the cardiovascular system, mammary glands, the intestine, and kidneys.⁴⁵ In the heart

(Figure 3), NRG-1 is expressed and released by the endocardial and microvasculature endothelium. NRG-1 binds to two of the four erythroblastic leukaemia viral oncogene homolog (ErbB) receptors, namely ErbB3 and ErbB4, of which ErbB4 seems to be the most important in the heart. Ligand binding activates these receptors and initiates dimerization, usually with ErbB2, although ErbB4 homodimer signalling has been described. Dimerization triggers the activation of signalling molecules, such as the mitogen-activated protein kinase (MAPK) and the phosphatidylinositol-3-OH kinase (PI3K)/ Akt pathways. Paracrine communication through NRG-1 between cardiac endothelial cells and cardiomycytes has been confirmed in co-culture studies,⁷ adding NRG-1 to a multiplicity of endothelial factors where interdependence contributes to the regulation of cardiovascular function.

In vitro effect of neuregulin-1 on cardiomyocytes and myocardial performance

Cultured cardiomyocytes exposed to NRG-1 show less cytochrome *c* release, caspase-3 activation, myofibrillar disorganization, and apoptotic cell death when exposed to oxidative stress and anthracyclines.^{46–48} Treatment of cultured adult rat ventricular myocytes with NRG- 1 induces DNA synthesis and cardiomyocyte proliferation through ErbB4 signalling.⁴⁹ In isolated cardiac muscle, NRG-1 reduced contractility and inotropic responsiveness to adrenergic stimulation. Most likely, this effect is linked to activation of eNOS; ErbB receptors and eNOS co-localize in the caveolae.⁵⁰ Interestingly, sympatholytic effects of NRG-1 have been recapitulated in the brain cardiovascular control system, where NRG-1 decreases heart rate and renal sympathetic nerve activity.^{51,52}

The beneficial effects of NRG-1 on cell viability, structure, and growth at the cardiomyocyte level, and the sympatholytic effects at the level of muscle and central nervous system offer an attractive therapeutic profile to NRG-1.

Endogenous neuregulin-1/erythroblastic leukaemia viral oncogene homolog signalling in cardiac disease

NRG-1/ErbB expression and signalling change during disease states. In compensatory states of ventricular hypertrophy, ErbB2 and ErbB4 expression increase, whereas depressed levels are found in terminal stages of HF.⁵³ Depression of NRG-1/ErbB signalling may contribute to progressive myocardial dysfunction. In addition, co-morbidities like diabetes may predispose to such depression.⁵⁴ Interestingly, in patients who had left-ventricular unloading with ventricular assist devices, depressed levels of ErbB receptors reversed.^{55,56}

Ischaemia/reperfusion injury is a potent activator of myocardial NRG-1/ErbB signalling.⁵⁷ Depressed endothelial NRG-1synthesis impairs recovery of cardiac contractile function after an ischaemic insult.⁵⁸

Cardiac therapeutic effect of neuregulin-1 in animals

The therapeutic effect of an EGF-domain fragment of rhNRG-1 β or a kringle and Ig domain-containing version (known as GGF2) was demonstrated in a series of animal models of HFREF. In rats with



Figure 3 NRG-1/ErbB signalling in the cardiomyocyte. NRG-1 regulates cardiomyocyte structure, mechanical performance, hypertrophic growth, proliferation, and viability. NRG-1, expressed as a transmembrane pro-protein is spliced by specific proteases, belonging to the desintegrin and metalloproteinase (ADAM) family and include ADAM 17 and ADAM 19. After binding to ErbB4, homodimerization of ErbB4 or heterodimerization with ErbB2, different signalling pathways are activated. These include the ErbB2/FAK/Src pathway, the activation of PI3K/Akt, the activation of ERK1/2, and the phosphorylation of eNOS, leading to an increase in NO production. This has been shown to attenuate adrenergic inotropic stimulation and phosphorylate phospholamban leading to increased SERCA2a activity and calcium uptake by the sarcoplasmic reticulum. Akt, protein kinase B; eNOS, endothelial nitric oxide synthase; ErbB2/4,erythroblastic leukaemia viral oncogene homolog 2/4; ERK1/2, extracellular signal-; PI3K, phosphoinositide-3-kinase; Src, proto-oncogene tyrosine-protein kinase.

systolic dysfunction induced by myocardial infarction, rhNRG-1 β administered daily for 5 days improved cardiac function.⁵⁹ Similar effects were reported in models of anthracycline and virally induced cardiac injury in mice. Next, in a model of diabetic cardioy-myopathy, rhNRG-1 improved LV function and reduced LV collagen volume fraction.⁶⁰

In large animals with rapid-pacing-induced HF, rhNRG-1 β similarly improved LV function. GGF2 treatment of rats or swine with myocardial infarction-induced myocardial dysfunction is associated with improved systolic function and reduced progressive remodelling.^{61,62}

Current clinically available interventions targeting endothelium-cardiomyocyte communication

The central hypothesis of this review is that generalized endothelial dysfunction is an important characteristic of chronic HF, and that alleviating endothelial dysfunction may improve HF symptoms and/or

prognosis. Medical interventions may restore endothelial function globally or partially, or may only substitute one of more endothelial communication pathways (Figure 4). For example, physical exercise may have a rather global, pleiotropic effect on endothelial function, whereas administration of a NO synthase agonist will be rather pathway specific. Since endothelial cells are present throughout the organism, endothelium-targeted interventions may improve the function of each organ in the organism. Retrospectively, it may be difficult to pinpoint how exactly this improvement emerged. Besides normalizing tissue perfusion or organ's haemodynamic loading conditions, the endothelium-targeted intervention may have enhanced a tissue-specific homeostatic process in the micro-capillaries, where endothelial cells similarly as in the heart directly interact with the organ's specific cells. Hence, a general principle in 'endothelium therapeutics' is to act gently and patiently, allowing endotheliumcontrolled homeostasis to recover, and not to shoot for abrupt changes of haemodynamic parameters like blood pressure or vascular resistance with excessive pharmacological dosages. Therapies directed at the NO-cGMP pathway may have a more direct and greater impact on haemodynamics than therapies directed at the NRG-1 pathway.



Figure 4 Cardiac endothelium-cardiomyocyte signalling pathways tested in clinical trials in 2014. Overview of currently tested drugs (and their action) targeting endothelium-cardiomyocyte crosstalk. As shown, these drugs act at different levels. For example, serelaxin increases the concentration of endothelial activators, and hence the endogenous release of endothelial factors, especially nitric oxide (NO). Neuregulin-1 (NRG-1)-like substances and NO donors mimic and thus replace endothelium-released factors. Soluble guanyl cyclase (sGC) and particulate guanyl cyclase (pGC) activators and phosphodiesterase-5 (PDE-5) inhibitors intensify the signalling cascade triggered by endothelial factors in the cardiomyocyte. Use of these drugs may be indicated in different phenotypes of patients, or at different stages of this disease. The combination of some drugs may be physiologically interesting, while other combinations may be dangerous given the risk to over-activate one of the pathways. GGF-2, Glial Growth Factor 2; EGF, epidermal growth factor; NEP neutral endogenous peptidase; cGMP, cyclic guanosine monophosphate; PKG, protein kinase G; ERK, extracellular-signal-regulated kinase.

Organic nitrates and nitric oxide donors

Therapeutic concentrations of organic nitrates preferentially vasodilate the veins and conduit arteries, lowering the peripheral arterial resistance only at higher doses. In the myocardium, NO has positive lusitropic effects with earlier onset of ventricular relaxation and improvement in ventricular compliance.^{15,63} Intravenous infusion of nitroprusside has been shown to improve ventricular distensibility and a reduction in LV peak systolic pressure.⁶⁴ The effects of NO on myocardial energetics are synergistic with its effects on cardiac contractility, by reducing myocardial energy wastage brought about by the myocardial contraction against reflected arterial pressure waves in late LV ejection.⁶⁵

The use of organic nitrates is not without limitations. Long-term use of organic nitrates is characterized by the development of tolerance. In addition, NO resistance occurs frequently in HF as a result of the oxidative stress. In the presence of oxidative stress, the shift of sGC redox equilibrium towards the oxidized form renders sGC unresponsive to NO. The abundance of superoxide anions in HF may lead to interaction with NO with formation of peroxynitrite. Peroxynitrite has numerous cytotoxic effects; through its interaction with lipids and proteins, it disrupts mitochondrial function and cellular signalling, eventually leading to cellular dysfunction and death. The addition of hydralazine to chronic nitrate therapy prevents nitrate tolerance and prevents the increase in vascular superoxide production.⁶⁶ In a small study of 28 HFREF patients, hydralazine has been shown to restore NO responsiveness.⁶⁷ Hence, the synergistic combination of hydralazine and organic nitrates is thought to exert beneficial effects on the endothelium.

Benefits of organic nitrates were first demonstrated in the Vasodilator Heart Failure Trial (V-HeFT I), where improvements in leftventricular function and survival were seen following the addition of isosorbide dinitrate and hydralazine to a HF therapeutic regimen of diuretics and digoxin.⁶⁸ The concept of augmenting NO bioavailability was subsequently tested in the African-American Heart Failure Trial (A-HeFT), where a fixed combination of isosorbide dinitrate and hydralazine was tested in a group of advanced HF patients.⁶⁹ This study was terminated prematurely due to benefits in the treatment arm, with improvements in mortality, HF admissions, and quality of life among those receiving isosorbide dinitrate and hydralazine in addition to standard HF therapy.

Phosphodiesterase-5 inhibition

Phosphodiesterases are a group of enzymes which catalyse cyclic nucleotide hydrolysis, thereby controlling the intracellular concentration of second messenger molecules including cAMP and cGMP.

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Phosphodiesterase-5 (PDE-5) inhibitors increase the intracellular concentration of cGMP, which leads to enhancement of endothelial function, 70 improvement of renal response to NPs, 71 and attenuation of adverse LV remodelling. 72

PDE-5 inhibitors have been explored in the management of pulmonary hypertension secondary to left-sided HF. In general, great caution should be exercised when using pulmonary-specific vasodilators in HF: selective pulmonary arterial vasodilatation without concomitant left-ventricular afterload reduction risks precipitating acute pulmonary oedema due to sudden increases in RV output emptying into a congested left atrium. This deleterious effect has been described with inhaled NO.^{73,74} Prostaglandins⁷⁵ and endothelin receptor antagonists⁷⁶ have been associated with progression of chronic HF. However, PDE-5 inhibitors have not been associated with this risk of worsening left HF. In fact, Guazzi et al.⁷⁷ demonstrated an improvement in exercise capacity and quality of life in small groups of patients with HFREF, as well as improvement in left- and RV function in HFPEF patients with pulmonary hypertension.⁷⁸ Unfortunately, these favourable findings were not seen in a multi-centre trial involving the use of sildenafil in 216 HFPEF patients (regardless of pulmonary hypertension).⁷⁹ There was no significant improvement in exercise capacity, quality of life, or haemodynamics. In addition, there was no significant increase in cGMP levels between the two groups. Failure to increase cGMP levels with PDE-5 inhibition suggests that the predominant problem may be decreased cGMP production, rather than increased cGMP breakdown by PDE-5.

Soluble guanyl cyclase stimulators and activators

Given that regulation of cardiovascular homeostasis by NO is mediated through sGC-cGMP-dependent downstream mechanisms, while the undesirable effects of NO appear independent of this signalling pathway, sGC has recently emerged as an attractive therapeutic target for HF. Two classes of compounds are known to influence the activity of sGC independent of NO, sGC stimulators and sGC activators.

sGC exists in two isoforms, with distinct sensitivity to NO, depending on the state of its prosthetic ferrous haem group. A reduced ferrous haem group is necessary for NO-dependent sGC stimulation. Conversely, oxidation of the haem group results in its dissociation, rendering the enzyme dysfunctional and resistant to NO stimulation.⁸⁰ sGC stimulators target the NO-sensitive, reduced sGC; they sensitize sGC to NO in the presence of low levels of NO and directly increase the activity of sGC in the absence of NO. In contrast, sGC activators specifically and effectively target NO-unresponsive sGC, irrespective of the bioavailability of endogenous NO.

sGC activators have potent vasodilatory effects, and have been shown to exert haemodynamic and other non-vascular effects in animal studies.^{81,82} Hypertensive rats treated with sGC activators demonstrated reductions in blood pressure and remodelling. In addition, sGC activators have renoprotective effects independent of their blood pressure lowering effects. In canine models of experimental HF, reductions in preload and afterload with concomitant improvements in renal perfusion and cardiac output were demonstrated following the administration of cinaciguat, a sGC activator.⁸³ These effects were dose-dependent. Similar beneficial effects were shown in rats with cardiorenal syndrome—there were attenuation of blood pressure, reduced fibrosis, and inflammation with improvement in cardiac and renal function, and reduced mortality.⁸⁴ Clinical trials involving HF patients have not replicated similar results. The efficacy and safety of intravenous cinaciguat was evaluated in a phase II study involving patients admitted for acute decompensated HF. This study was discontinued prematurely due to excess hypotension in the patients.⁸⁵ (see below recommendation 3 under 'Considerations for future clinical trials in HF').

sGC stimulators are potent vasodilators, exerting similar favourable effects on blood pressure, LV remodelling, and cardio- and renoprotective effects.^{86,87} Experiments involving hypertensive rats demonstrated lower myocardial remodelling, attenuated cardiac, and renal fibrosis with improvement in renal function, following administration of riociguat.⁸⁸ In canine models of HF, BAY 41-2272 resulted in improvements in systemic vasoconstriction without significant effects on blood pressure, pulmonary hypertension, cardiac output, and renal function.⁸⁹ The oral sGC stimulator BAY 60-4552 was evaluated in patients with HFREF and secondary pulmonary hypertension, and demonstrated reductions in left- and RV preload and afterload.⁹⁰ The Acute Hemodynamic Effects of Riociguat in Patients with Pulmonary Hypertension Associated with Diastolic Heart Failure (DILATE-1) study was a phase IIa trial evaluating the oral sGC stimulator riociguat in HFPEF patients with pulmonary hypertension. Following a single dose of riociguat 2 mg, there were significant increases in stroke volume and cardiac index, with concomitant decreases in afterload and RV area.⁹¹ Other phase II and III trials involving the use of sGC stimulators in HF are currently ongoing. The Soluble Guanylate Cyclase stimulator Heart Failure Studies (SOCRATES) program consists of two parallelgroup, randomized, double-blind, multi-centre phase II studies evaluating the pharmacodynamics, pharmacokinetics, safety, and tolerability of the new oral sGC stimulator BAY1021189 in patients with worsening HF requiring hospitalization (ClinicalTrials.gov Identifier: NCT101951638). Eligible patients will be recruited into either SOCRATES-REDUCED (patients with LVEF <45%) or SOCRATES-PRESERVED (patients with LVEF \geq 45%), where four dose regimens of this drug will be explored over 12 weeks.

Neprilysin inhibitors

LCZ 696 is a complex molecule which comprises the neprilysin inhibitor prodrug AHU377 together with an angiotensin II receptor blocker (ARB) valsartan in the same compound.⁹² Inhibition of neprilysin enhances the availability of biologically active NPs. The latter peptides act in parallel to the NO-sGC-cGMP pathway via activation of the NP-pCG-cGMP signalling system. NPs enhance natriuresis, diuresis, and vasodilatation. They also augment myocardial relaxation through a cardiac endothelium-dependent signalling pathway⁹³ and reduce myocardial hypertrophy through increased generation of cGMP.^{94,95} Inhibition of angiotensin receptor results in suppression of the renin–angiotensin–aldosterone system, which has been proved to be beneficial in HF.

LCZ 696 has been successfully evaluated in HFREF patients. The PARADIGM-HF (Prospective comparison of ARNI with ACEI (angiotensin-converting enzyme inhibition) to Determine Impact on Global Mortality and morbidity in Heart Failure) study is a phase III, randomized, double-blind, parallel group, two-arm, event-driven trial evaluating the safety and efficacy of LCZ 696 and enalapril in chronic symptomatic HFREF patients.⁹⁶ This trial has recently been completed and published; LCZ 696 resulted in a 16% reduction in all-cause mortality, 20% reduction in cardiovascular mortality and 21% reduction in HF hospitalizations among chronic HFREF patients.⁹⁷ Although augmentation of the NP pathway by LCZ 696 may be expected to boost endothelial function, other beneficial neurohormones may also be involved and endothelial function was not specifically measured in this landmark trial. Future studies in this cohort may provide insight to the extent to which beneficial effects of LCZ696 may be attributable to improvement in endothelial function.

LCZ696 was also evaluated in the PARAMOUNT [Prospective comparison of ARNI (angiotensin receptor neprilysin inhibitor) with ARB on Management Of heart failUre with preserved ejection fracTion] trial; it was a phase II, randomized, parallel-group, multicentre trial comparing LCZ 696 against valsartan over 36 weeks in 301 HFPEF patients.⁹⁸ LCZ 696 reduced N-terminal prohormone of brain natriuretic peptide (NT proBNP) to a greater extent than valsartan at 12 weeks, but the difference was not significant at 36 weeks. Favourable changes in left-atrial volume and dimensions were observed at 36 weeks, without concomitant changes in diastolic function. A large phase III trial has recently commenced to evaluate the benefit of LCZ 696 compared with Valsartan, on Morbidity and Mortality in Heart Failure Patients With Preserved Ejection Fraction (PARAGON-HF), ClinicalTrials.gov Identifier: NCT01920711].

Serelaxin

Serelaxin is the recombinant form of human relaxin-2, a naturally occurring peptide hormone that mediates systemic haemodynamic and renal adaptive changes during pregnancy.⁹⁹ Serelaxin binds to relaxin family peptide (RXFP) receptors that are located in the heart, blood vessels,¹⁰⁰ and kidneys. Serelaxin reduces pulmonary capillary wedge pressure and systemic vascular resistance, and increases cardiac index in stable and acute HF patients.^{101,102} These haemodynamic effects of serelaxin are predominantly a consequence of receptormediated actions on endothelial cells, including the release of NO, a reduction of endothelin-1 levels, and an increase in vascular endothelial growth factors.^{103–107} Apart from these endothelial effects, promotes cell survival, and mitigates reparative fibrosis.¹⁰⁸

In the phase III RELAX-AHF^{101,102} trial in patients with acute HF, a 48 h infusion of serelaxin significantly reduced symptoms and signs of congestion and lowered NT-proBNP levels compared with placebo. It also shortened length of hospital stay. Interestingly, at Day 180, serelaxin reduced all-cause mortality and cardiovascular death, although these data were not primary endpoints of the trial and need to be confirmed in an adequately powered prospective trial.

Exercise

Exercise enhances endothelial function, in particular NO bioavailability, and it improves NO-mediated vasodilation in chronic HF¹⁰⁹ primarily through increased endothelial shear stress. In addition, activation of various interconnected endothelial pathways, resulting from altered plasma levels of circulating insulin, glucose, lipids, interleukins, all known to affect endothelial (dys)function, may also contribute to the beneficial effects of exercise. *In vivo* studies have demonstrated an upregulation of vascular eNOS expression in humans and animals exposed to physical training.¹¹⁰ In addition, endothelial shear stress promotes NO release through the increase in Ca²⁺ influx¹¹¹ and activation of endothelial K⁺ channels.¹¹² This improvement in NO-mediated vasodilatation is associated with improvements in peak oxygen consumption, indicating that an improvement in endothelial function enhances cardiorespiratory fitness.⁴¹

However, these favourable findings were not replicated in the Heart Failure: A Controlled Trial Investigating Outcomes of Exercise Training (HF-ACTION) study, a multi-centre, randomizedcontrolled trial evaluating the efficacy of home-based exercise training following a period of supervised exercise training in chronic HFREF patients.¹¹³ However, the non-adherence rate of 40% in the exercise arm and crossovers to exercise in the control arm could have contributed, in part, to the neutral findings. Endothelial function was neither a selection criterion nor an assessment parameter, and it is conceivable that a study sample enriched with endothelial dysfunction may have demonstrated benefits of exercise training. Supporting this postulation, a sub-analysis of women in HF-ACTION, in whom age-related endothelial dysfunction has been shown to be more prevalent than in men,¹¹⁴ showed that exercise training in women with HF was associated with a larger reduction in rate of the combined endpoint of all-cause mortality and hospital stay than in men.¹¹⁵

The benefits of exercise training are similarly demonstrated in HFPEF, where exercise training has been shown to be associated with improvements in peak oxygen consumption, physical functioning score, and echocardiographic indices of diastolic function.^{116,117} However, a study performed by Kitzman et al.¹¹⁸ showed that neither carotid arterial stiffness nor brachial artery flow-mediated vasodilatation accounted for improvements in peak oxygen consumption seen with exercise training. The duration of endurance training may not have been sufficient to produce detectable changes in conduit arterial function, and potential benefits on the microvasculature mediated by NO-dependent vasodilatation, cannot be excluded. Indeed, Borlaug et al.⁴² showed that microvascular reserve, assessed by change in EndoPAT fingertip plethysmography in response to exercise, was reduced in patients with HFPEF compared with age-matched healthy controls, and both systemic vascular conductance and microvascular reserve were related to peak VO2 in HFPEF. Thus, although conduit arterial endothelial function seems to be preserved in HFPEF, impaired microvascular function may limit exercise performance in older patients with HFPEF by decreasing diffusive oxygen transport to the active muscle.

Human recombinant neuregulin-1

The effects of recombinant human NRG-1 (rhNRG-1) in patients with stable HFREF on optimal medical therapy have been described in two recent phase II clinical trials.^{119,120} A first trial, in which a daily intravenous infusion of rhNRG-1 was given to patients during 10 consecutive days, showed an increase in left-ventricular performance and reverse LV remodelling 30 days later. On Day 90, LV function improved even further and LV dimensions further decreased.¹¹⁹ A second trial, in which patients received a consecutive 12 h intravenous infusion of rhNRG-1 during 10 days, showed a 30% increase in cardiac output and a 20–30% decrease in pulmonary artery wedge

pressure and systemic vascular resistance of 30%. Twelve weeks after infusion, LVEF was increased by 12%.¹²⁰ Importantly, in these studies, short-term rhNRG-1 administration was safe and well tolerated. Larger trials are now ongoing. A phase II clinical trial has been set out to determine, over a period of 12 months, the efficacy and safety of NRG-1 as a treatment in 120 patients with chronic HFREF (ClinicalTrials.gov Identifier: NCT01251406), and a phase III clinical trial has been set-up with the purpose of evaluating the ability of NRG-1 to improve survival (ClinicalTrials.gov Identifier: NCT01541202), cardiac remodelling (ClinicalTrials.gov Identifier: NCT01439893), and efficacy and safety of subcutaneous NRG-1 administration in HFREF (ClinicalTrials.gov Identifier: NCT01214096). A Phase I clinical trial with Glial growth factor 2, an NRG-1 isoform, is also trying to determine its effects on patients with LV dysfunction and symptomatic HF (ClinicalTrials.gov Identifier: NCT01258387).

A potential risk of systemically treating patients with NRG-1 is the activation of neoplastic signalling and cancer growth through the induction of ErbB3/ErbB2 oncogenic complex in ErbB2 amplified cells. To overcome this risk, Lee *et al.*¹²¹ designed an engineered bivalent neuregulin-1 (NN) that demonstrates reduced neoplastic potential in comparison with NRG-1, and still retains its cardioprotective properties. Although its clinical effects still have to be demonstrated, NN could represent a more translationally relevant therapy to rescue ErbB signalling in chronic diseases such as HF.

Considerations for future clinical trials in heart failure

The HF clinical trial landscape, and particularly that of HFPEF, is littered with 'gravestones' of recent neutral trials. Some of these have included trials of agents known to have positive effects on endothelial function, such as ACE inhibitors and ARBs in HFPEF (PEP-CHF, CHARM Preserved, I-PRESERVE). However, endothelial dysfunction was neither a selection criterion nor specifically assessed in these prior trials. Hence, not the medical action of the drugs, but the design of the clinical trials may have contributed to the failure of many of these HF trials. The design of a trial (e.g. patient selection and study endpoints) should be better adapted to both the action and the clinical expectations of the drug of interest. In the further clinical development of tools to target endothelial signalling in HF, we recommend the following general principles (*Figure 5*):

First, patient selection may be based on phenotypic characteristics associated with endothelial dysfunction (e.g. metabolic dysfunction, obesity, diabetes, males vs. females), rather than solely on vague traditional HF parameters like LVEF or stand-alone biomarkers; in other words future trials may be designed to be more pathophysiologyspecific. This approach will obviously significantly scale down the number of patient candidates for drug development, but this may be better than vague studies in a broad, heterogeneous (and commercially more interesting) HF population. Although we believe that phenotypic characteristics of a patient more conveniently reflect patient-specific disease processes than any stand-alone biomarker or HF parameter, direct assessment of endothelial function in endothelium-targeted HF trials would be most logical. Such assessments will further homogenize and characterize the HF study population in terms of endothelial features, and allow longitudinal follow-up of endothelial function during the course of the clinical trial. However, accurate functional assessment of the endothelial system is complex, and functional testing of the endothelium should not be confined to a mere assessment of endothelial regulation of vascular motricity.

To date, most currently available indices of endothelial function have scientific downsides and it is not clear which of these indices is to be preferred. Some of the indices are often difficult to obtain, have not been rigorously standardized or show large intra- and interobserver variability. Notably, they merely reflect a narrow window of endothelial function such as NO-dependent effects or inflammatory changes, or lack specificity for endothelial dysfunction. Endothelial assessments applied to HF include (i) invasive or non-invasive measures of vasomotor function, (ii) serum markers of endothelial (dys)function such as endothelin-1, NRG-1, C-reactive protein, von Willebrand factor, E-selectin, thrombospondin-1, endothelial miRNAs (miR-146a), and endothelial microparticles, (iii) urine markers of endothelial dysfunction such as albuminuria. or (iv) circulating cells such as endothelial progenitor cells. A recent study involving community-based adults without cardiovascular diseases demonstrated associations between endothelial microparticles and cardiometabolic risk factors, highlighting the importance of such risk factors on endothelial function.¹²² Other markers of endothelial function have shown correlations with adverse events or prognosis of HF.^{123,124} Yet, their use in clinical cardiology or in scientific trials is still very limited. Obviously, further progress and brilliant discoveries in the assessment of endothelial function in HF are awaited.

Second, intermediate endpoints such as quality-of-life indices, exercise capacity, and some biomarkers of matrix remodelling and anatomic cardiac indices may be considered, as these may be useful surrogates for HF mitigating drug activities, induced by these 'endothelial' interventions. Two recent studies illustrated this, showing anti-fibrotic effects of NRG-1¹²⁵ and LCZ696 in the remodelling myocardium.¹²⁶ These intermediate endpoints may be coupled with biomarkers of HF prognosis, such as serum BNP, LV diameters, left-atrial size, or heart-rate variability. Too many active HF drugs have been terminated based on their disappointing effect on total mortality, despite a beneficial effect on the progression of chronic HF. This is unfortunate since in HF, e.g. for the very symptomatic and elderly patients, drugs can be useful even if they do not reduce total mortality.

Finally, preference should be given to pleiotropic drugs acting at various sites, including the endothelium, in order to reestablish overall physiological homeostasis. For drugs acting through endothelial-mediated signalling pathways, this would necessitate prolonged exposure to the lowest possible dosages even if at first no measurable changes are observed. For example, as half-life and duration of signalling activity varies significantly among the different interdependent endothelium-cardiomyocyte signalling pathways, one may expect a profound mismatch in their interaction in HF. This mismatch may be further hampered if only one of these pathways would be targeted for short periods and at high dosages of a drug. In contrast, chronic exposure to lower dosages would, i.p. in HF patients with concomitant pulmonary hypertension, allow for an autoamplifying shear stress-induced pulmonary endothelial-mediated haemodynamic reversal back into a low impedance pulmonary vascular system. Similarly, in the systemic circulation, as most of



Figure 5 Designing clinical trials to test endothelium–cardiomyocyte pathways in heart failure. When a drug is tested in a clinical trial there should be evidence that the drug-related disease pathway is disturbed in the study population. Therefore, when 'endothelial' drugs are put into trial, one should first carefully define a study population in which cardiac endothelial dysfunction is central in the pathogenesis. One way to do this is to start from phenotypic characteristics and co-morbidities known to reflect or to be associated with endothelial dysfunction (independent from LVEF). Selecting endpoints closer to the presumed cardiac effect of the drug (e.g. markers of reverse cardiac and matrix remodelling, and of diastolic dysfunction) would allow a cheaper 'check-and-balance' strategy. Also, it may suffice to define the patient population positively responding to the trial drug. If required, the results could then later be employed to test effects on mortality and hospitalization in a pathophysiological coherent and drug-responsive population.

these drugs are vasodilators, focus in HF should be on the modulation of arterial impedance while avoiding arterial pressure drop. A prolonged exposure at low dosages of endothelium-mediated drugs would thus be expected to optimize the ventricular-arterial crosstalk through simultaneous endothelium-mediated effects on timing and rate of ventricular relaxation and on arterial impedance while avoiding arterial pressure drop. In summary, preference may be given to low and prolonged dosages of pleiotropic drugs, while avoiding short treatments at high dosages of single target drugs which may be detrimental to endothelial-cardiomycyte balance. In addition, this low-dosage pleiotropic drug strategy would greatly complement the more physiological, but still largely undervalued exercise training.

Accordingly, a phenotype-based check-and-balance approach may allow a stepwise progression of new clinical tools in cheaper clinical studies to define which patients are optimal candidates for treatment.

Conclusions

Endothelium-controlled signalling pathways play a crucial homeostatic role in cardiac tissue. Disturbance of these pathways accounts for the adverse myocardial remodelling and dysfunction seen in HF, and portends a worse prognosis in these patients. Endothelial pathways are emerging as an important pathophysiological target in the treatment of HF regardless of EF. Attention should be paid to patient selection and study endpoints in future clinical trials, in order to avoid new disappointments in the clinical development of this target.

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References

- Aranda JM Jr, Johnson JW, Conti JB. Current trends in heart failure readmission rates: analysis of Medicare data. *Clin Cardiol* 2009;32:47–52.
- Solomon SD, Dobson J, Pocock S, Skali H, McMurray JJ, Granger CB, Yusuf S, Swedberg K, Young JB, Michelson EL, Pfeffer MA, Candesartan in Heart failure: Assessment of Reduction in Mortality and morbidity (CHARM) Investigators. Influence of nonfatal hospitalization for heart failure on subsequent mortality in patients with chronic heart failure. *Circulation* 2007;**116**:1482–1487.
- Levy D, Kenchaiah S, Larson MG, Benjamin EJ, Kupka MJ, Ho KK, Murabito JM, Vasan RS. Long-term trends in the incidence of and survival with heart failure. N Engl J Med 2002;347:1397–1402.
- Roger VL, Weston SA, Redfield MM, Hellermann-Homan JP, Killian J, Yawn BP, Jacobsen SJ. Trends in heart failure incidence and survival in a community-based population. JAMA 2004;292:344–350.
- Lam CS, Donal E, Kraigher-Krainer E, Vasan RS. Epidemiology and clinical course of heart failure with preserved ejection fraction. *Eur J Heart Fail* 2011;**13**:18–28.
- Tirziu D, Giordano FJ, Simons M. Cell communications in the heart. *Circulation* 2010; 122:928–937.

- 7. Brutsaert DL. Cardiac endothelial-myocardial signaling: its role in cardiac growth, contractile performance, and rhythmicity. *Physiol Rev* 2003;**83**:59–115.
- Brutsaert DL. Role of endocardium in cardiac overloading and failure. Eur Heart J 1990;11(Suppl. G):8–16.
- Brutsaert DL, Meulemans AL, Sipido KR, Sys SU. Effects of damaging the endocardial surface on the mechanical performance of isolated cardiac muscle. *Circ Res* 1988;62:358–366.
- Li K, Rouleau JL, Andries LJ, Brutsaert DL. Effect of dysfunctional vascular endothelium on myocardial performance in isolated papillary muscles. *Circ Res* 1993;72: 768–777.
- Pinsky DJ, Patton S, Mesaros S, Brovkovych V, Kubaszewski E, Grunfeld S, Malinski T. Mechanical transduction of nitric oxide synthesis in the beating heart. *Circ Res* 1997;81:372–379.
- Castro LR, Verde I, Cooper DM, Fischmeister R. Cyclic guanosine monophosphate compartmentation in rat cardiac myocytes. *Circulation* 2006;113:2221–2228.
- Takimoto E, Belardi D, Tocchetti CG, Vahebi S, Cormaci G, Ketner EA, Moens AL, Champion HC, Kass DA. Compartmentalization of cardiac beta-adrenergic inotropy modulation by phosphodiesterase type 5. *Circulation* 2007;**115**:2159–2167.
- Fischmeister R, Castro LR, Abi-Gerges A, Rochais F, Jurevicius J, Leroy J, Vandecasteele G. Compartmentation of cyclic nucleotide signaling in the heart: the role of cyclic nucleotide phosphodiesterases. *Circ Res* 2006;**99**:816–828.
- Mohan P, Brutsaert DL, Paulus WJ, Sys SU. Myocardial contractile response to nitric oxide and cGMP. *Circulation* 1996;93:1223–1229.
- Rastaldo R, Pagliaro P, Cappello S, Penna C, Mancardi D, Westerhof N, Losano G. Nitric oxide and cardiac function. *Life Sci* 2007;81:779–793.
- Vila-Petroff MG, Younes A, Egan J, Lakatta EG, Sollott SJ. Activation of distinct cAMP-dependent and cGMP-dependent pathways by nitric oxide in cardiac myocytes. *Circ Res* 1999;84:1020–1031.
- Kojda G, Kottenberg K. Regulation of basal myocardial function by NO. Cardiovasc Res 1999;41:514–523.
- Layland J, Li JM, Shah AM. Role of cyclic GMP-dependent protein kinase in the contractile response to exogenous nitric oxide in rat cardiac myocytes. *J Physiol* 2002; 540(Pt 2):457–467.
- LeWinter MM, Granzier H. Cardiac titin: a multifunctional giant. *Circulation* 2010; 121:2137–2145.
- Brutsaert DL. Cardiac dysfunction in heart failure: the cardiologist's love affair with time. Prog Cardiovasc Dis 2006;49:157–181.
- Brutsaert DL, Sys SU. Relaxation and diastole of the heart. *Physiol Rev* 1989;69: 1228–1315.
- Musialek P, Lei M, Brown HF, Paterson DJ, Casadei B. Nitric oxide can increase heart rate by stimulating the hyperpolarization-activated inward current, I(f). *Circ Res* 1997;81:60–68.
- Balligand JL, Kelly RA, Marsden PA, Smith TW, Michel T. Control of cardiac muscle cell function by an endogenous nitric oxide signaling system. *Proc Natl Acad Sci USA* 1993;**90**:347–351.
- Recchia FA, McConnell PI, Bernstein RD, Vogel TR, Xu X, Hintze TH. Reduced nitric oxide production and altered myocardial metabolism during the decompensation of pacing-induced heart failure in the conscious dog. *Circ Res* 1998;83: 969–979.
- Scherrer-Crosbie M, Ullrich R, Bloch KD, Nakajima H, Nasseri B, Aretz HT, Lindsey ML, Vancon AC, Huang PL, Lee RT, Zapol WM, Picard MH. Endothelial nitric oxide synthase limits left ventricular remodeling after myocardial infarction in mice. *Circulation* 2001;**104**:1286–1291.
- Paulus WJ, Tschope C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. J Am Coll Cardiol 2013;62:263–271.
- Schulz E, Jansen T, Wenzel P, Daiber A, Munzel T. Nitric oxide, tetrahydrobiopterin, oxidative stress, and endothelial dysfunction in hypertension. *Antioxid Redox Signal* 2008;10:1115–1126.
- van Heerebeek L, Hamdani N, Falcao-Pires I, Leite-Moreira AF, Begieneman MP, Bronzwaer JG, van der Velden J, Stienen GJ, Laarman GJ, Somsen A, Verheugt FW, Niessen HW, Paulus WJ. Low myocardial protein kinase G activity in heart failure with preserved ejection fraction. *Circulation* 2012;**126**:830–839.
- Matsubara BB, Matsubara LS, Zornoff LA, Franco M, Janicki JS. Left ventricular adaptation to chronic pressure overload induced by inhibition of nitric oxide synthase in rats. *Basic Res Cardiol* 1998;93:173–181.
- Bishu K, Hamdani N, Mohammed SF, Kruger M, Ohtani T, Ogut O, Brozovich FV, Burnett JC Jr, Linke WA, Redfield MM. Sildenafil and B-type natriuretic peptide acutely phosphorylate titin and improve diastolic distensibility in vivo. *Circulation* 2011;**124**:2882–2891.
- Cooper CJ, Landzberg MJ, Anderson TJ, Charbonneau F, Creager MA, Ganz P, Selwyn AP. Role of nitric oxide in the local regulation of pulmonary vascular resistance in humans. *Circulation* 1996;93:266–271.

- Moraes DL, Colucci WS, Givertz MM. Secondary pulmonary hypertension in chronic heart failure: the role of the endothelium in pathophysiology and management. *Circulation* 2000;**102**:1718–1723.
- Bock JS, Gottlieb SS. Cardiorenal syndrome: new perspectives. *Circulation* 2010; 121:2592–2600.
- Zeiher AM, Krause T, Schächinger V, Minners J, Moser E. Impaired Endothelium-Dependent Vasodilation of Coronary Resistance Vessels Is Associated With Exercise-Induced Myocardial Ischemia. *Circulation* 1995;91:2345–2352.
- 36. Gheorghiade M, Marti CN, Sabbah HN, Roessig L, Greene SJ, Bohm M, Burnett JC, Campia U, Cleland JG, Collins SP, Fonarow GC, Levy PD, Metra M, Pitt B, Ponikowski P, Sato N, Voors AA, Stasch JP, Butler J, Academic Research Team in Heart F. Soluble guanylate cyclase: a potential therapeutic target for heart failure. *Heart Fail Rev* 2013;**18**:123–134.
- Katz SD, Biasucci L, Sabba C, Strom JA, Jondeau G, Galvao M, Solomon S, Nikolic SD, Forman R, LeJemtel TH. Impaired endothelium-mediated vasodilation in the peripheral vasculature of patients with congestive heart failure. J Am Coll Cardiol 1992;19:918–925.
- Kubo SH, Rector TS, Bank AJ, Williams RE, Heifetz SM. Endothelium-dependent vasodilation is attenuated in patients with heart failure. *Circulation* 1991;84: 1589–1596.
- Drexler H, Zeiher AM, Meinzer K, Just H. Correction of endothelial dysfunction in coronary microcirculation of hypercholesterolaemic patients by L-arginine. *Lancet* 1991;**338**:1546–1550.
- Fischer D, Rossa S, Landmesser U, Spiekermann S, Engberding N, Hornig B, Drexler H. Endothelial dysfunction in patients with chronic heart failure is independently associated with increased incidence of hospitalization, cardiac transplantation, or death. *Eur Heart J* 2005;26:65–69.
- Hambrecht R, Fiehn E, Weigl C, Gielen S, Hamann C, Kaiser R, Yu J, Adams V, Niebauer J, Schuler G. Regular physical exercise corrects endothelial dysfunction and improves exercise capacity in patients with chronic heart failure. *Circulation* 1998;98:2709–2715.
- Borlaug BA, Olson TP, Lam CS, Flood KS, Lerman A, Johnson BD, Redfield MM. Global cardiovascular reserve dysfunction in heart failure with preserved ejection fraction. J Am Coll Cardiol 2010;56:845–854.
- Farrero M, Blanco I, Batlle M, Santiago E, Cardona M, Vidal B, Castel MA, Sitges M, Barbera JA, Perez-Villa F. Pulmonary hypertension is related to peripheral endothelial dysfunction in heart failure with preserved ejection fraction. *Circ Heart Fail* 2014; 7:791–798.
- 44. Akiyama E, Sugiyama S, Matsuzawa Y, Konishi M, Suzuki H, Nozaki T, Ohba K, Matsubara J, Maeda H, Horibata Y, Sakamoto K, Sugamura K, Yamamuro M, Sumida H, Kaikita K, Iwashita S, Matsui K, Kimura K, Umemura S, Ogawa H. Incremental prognostic significance of peripheral endothelial dysfunction in patients with heart failure with normal left ventricular ejection fraction. J Am Coll Cardiol 2012;60:1778–1786.
- Britsch S. The neuregulin-I/ErbB signaling system in development and disease. Adv Anat Embryol Cell Biol 2007;190:1–65.
- Kuramochi Y, Cote GM, Guo X, Lebrasseur NK, Cui L, Liao R, Sawyer DB. Cardiac endothelial cells regulate reactive oxygen species-induced cardiomyocyte apoptosis through neuregulin-1β/erbB4 signaling. J Biol Chem 2004;279:51141–51147.
- Kuramochi Y, Lim CC, Guo X, Colucci WS, Liao R, Sawyer DB. Myocyte contractile activity modulates norepinephrine cytotoxicity and survival effects of neuregulin-1 beta. Am J Physiol Cell Physiol 2004;286:C222–C229.
- Fukazawa R, Miller TA, Kuramochi Y, Frantz S, Kim YD, Marchionni MA, Kelly RA, Sawyer DB. Neuregulin-1 protects ventricular myocytes from anthracyclineinduced apoptosis via erbB4-dependent activation of PI3-kinase/Akt. J Mol Cell Cardiol 2003;35:1473–1479.
- Bersell K, Arab S, Haring B, Kuhn B. Neuregulin1/ErbB4 signaling induces cardiomyocyte proliferation and repair of heart injury. *Cell* 2009;**138**:257–270.
- Zhao YY, Feron O, Dessy C, Han X, Marchionni MA, Kelly RA. Neuregulin signaling in the heart. Dynamic targeting of erbB4 to caveolar microdomains in cardiac myocytes. *Circ Res* 1999;84:1380–1387.
- Matsukawa R, Hirooka Y, Ito K, Sunagawa K. Inhibition of neuregulin-1/ErbB signaling in the rostral ventrolateral medulla leads to hypertension through reduced nitric oxide synthesis. Am J Hypertens 2013;26:51–57.
- Matsukawa R, Hirooka Y, Nishihara M, Ito K, Sunagawa K. Neuregulin-1/ErbB signaling in rostral ventrolateral medulla is involved in blood pressure regulation as an antihypertensive system. J Hypertens 2011;29:1735–1742.
- Lemmens K, Doggen K, De Keulenaer GW. Role of neuregulin-1/ErbB signaling in 2cardiovascular physiology and disease: implications for therapy of heart failure. *Circulation* 2007;**116**:954–960.
- Gui C, Zhu L, Hu M, Lei L, Long Q. Neuregulin-1/ErbB signaling is impaired in the rat model of diabetic cardiomyopathy. *Cardiovasc Pathol* 2012;21:414–420.
- Rohrbach S, Niemann B, Silber RE, Holtz J. Neuregulin receptors erbB2 and erbB4 in failing human myocardium – depressed expression and attenuated activation. *Basic Res Cardiol* 2005;100:240–249.

- Uray IP, Connelly JH, Thomazy V, Shipley GL, Vaughn WK, Frazier OH, Taegtmeyer H, Davies PJ. Left ventricular unloading alters receptor tyrosine kinase expression in the failing human heart. J Heart Lung Transplant 2002;21: 771–782.
- Iivanainen E, Paatero I, Heikkinen SM, Junttila TT, Cao R, Klint P, Jaakkola PM, Cao Y, Elenius K. Intra- and extracellular signaling by endothelial neuregulin-1. *Exp Cell Res* 2007;**313**:2896–2909.
- Hedhli N, Huang Q, Kalinowski A, Palmeri M, Hu X, Russell RR, Russell KS. Endothelium-derived neuregulin protects the heart against ischemic injury. *Circulation* 2011;**123**:2254–2262.
- Liu X, Gu X, Li Z, Li X, Li H, Chang J, Chen P, Jin J, Xi B, Chen D, Lai D, Graham RM, Zhou M. Neuregulin-1/erbB-activation improves cardiac function and survival in models of ischemic, dilated, and viral cardiomyopathy. J Am Coll Cardiol 2006;48: 1438–1447.
- Li B, Zheng Z, Wei Y, Wang M, Peng J, Kang T, Huang X, Xiao J, Li Y, Li Z. Therapeutic effects of neuregulin-1 in diabetic cardiomyopathy rats. *Cardiovasc Diabetol* 2011; 10:69.
- Kasasbeh E, Murphy A, Williams P, Nunnally A, Adcock J, Caggiano A, Parry T, Ganguly A, Sawyer DB, Cleator JH. Neuregulin-1β improves cardiac remodeling after myocardial infarction in swine. *Circulation* 2011;**124**:A15531.
- Guo YF, Zhang XX, Liu Y, Duan HY, Jie BZ, Wu XS. Neuregulin-1 attenuates mitochondrial dysfunction in a rat model of heart failure. *Chin Med J (Engl)* 2012;**125**: 807–814.
- Shah AM, Spurgeon HA, Sollott SJ, Talo A, Lakatta EG. 8-bromo-cGMP reduces the myofilament response to Ca2+ in intact cardiac myocytes. *Circ Res* 1994;**74**: 970–978.
- Paulus WJ, Vantrimpont PJ, Shah AM. Acute effects of nitric oxide on left ventricular relaxation and diastolic distensibility in humans. Assessment by bicoronary sodium nitroprusside infusion. *Circulation* 1994;89:2070–2078.
- Paulus WJ, Bronzwaer JG. Nitric oxide's role in the heart: control of beating or breathing? Am J Physiol Heart Circ Physiol 2004;287:H8–13.
- 66. Daiber A, Oelze M, Coldewey M, Kaiser K, Huth C, Schildknecht S, Bachschmid M, Nazirisadeh Y, Ullrich V, Mulsch A, Munzel T, Tsilimingas N. Hydralazine is a powerful inhibitor of peroxynitrite formation as a possible explanation for its beneficial effects on prognosis in patients with congestive heart failure. *Biochem Biophys Res Commun* 2005;**338**:1865–1874.
- Gogia H, Mehra A, Parikh S, Raman M, Ajit-Uppal J, Johnson JV, Elkayam U. Prevention of tolerance to hemodynamic effects of nitrates with concomitant use of hydralazine in patients with chronic heart failure. J Am Coll Cardiol 1995;26: 1575–1580.
- 68. Cohn JN, Archibald DG, Ziesche S, Franciosa JA, Harston WE, Tristani FE, Dunkman WB, Jacobs W, Francis GS, Flohr KH, Goldman S, Cobb FR, Shah PM, Saunders R, Fletcher RD, Loeb HS, Hughes VC, Baker B. Effect of vasodilator therapy on mortality in chronic congestive heart failure. Results of a Veterans Administration Cooperative Study. N Engl J Med 1986;**314**:1547–1552.
- Taylor AL, Ziesche S, Yancy C, Carson P, D'Agostino R Jr, Ferdinand K, Taylor M, Adams K, Sabolinski M, Worcel M, Cohn JN, African-American Heart Failure Trial I. Combination of isosorbide dinitrate and hydralazine in blacks with heart failure. N Engl J Med 2004;351:2049–2057.
- Katz SD, Balidemaj K, Homma S, Wu H, Wang J, Maybaum S. Acute type 5 phosphodiesterase inhibition with sildenafil enhances flow-mediated vasodilation in patients with chronic heart failure. J Am Coll Cardiol 2000;36:845–851.
- 71. Chen HH, Huntley BK, Schirger JA, Cataliotti A, Burnett JC Jr. Maximizing the renal cyclic 3'-5'-guanosine monophosphate system with type V phosphodiesterase inhibition and exogenous natriuretic peptide: a novel strategy to improve renal function in experimental overt heart failure. J Am Soc Nephrol 2006;**17**:2742–2747.
- Takimoto E, Champion HC, Li M, Belardi D, Ren S, Rodriguez ER, Bedja D, Gabrielson KL, Wang Y, Kass DA. Chronic inhibition of cyclic GMP phosphodiesterase 5A prevents and reverses cardiac hypertrophy. *Nat Med* 2005;11: 214–222.
- Boilson BA, Schirger JA, Borlaug BA. Caveat medicus! Pulmonary hypertension in the elderly: a word of caution. *Eur J Heart Fail* 2010;**12**:89–93.
- Semigran MJ, Cockrill BA, Kacmarek R, Thompson BT, Zapol WM, Dec GW, Fifer MA. Hemodynamic effects of inhaled nitric oxide in heart failure. J Am Coll Cardiol 1994;24:982–988.
- Califf RM, Adams KF, McKenna WJ, Gheorghiade M, Uretsky BF, McNulty SE, Darius H, Schulman K, Zannad F, Handberg-Thurmond E, Harrell FE Jr., Wheeler W, Soler-Soler J, Swedberg K. A randomized controlled trial of epoprostenol therapy for severe congestive heart failure: The Flolan International Randomized Survival Trial (FIRST). Am Heart J 1997;**134**:44–54.
- Anand I, McMurray J, Cohn JN, Konstam MA, Notter T, Quitzau K, Ruschitzka F, Luscher TF. Long-term effects of darusentan on left-ventricular remodelling and clinical outcomes in the EndothelinA Receptor Antagonist Trial in Heart Failure (EARTH): randomised, double-blind, placebo-controlled trial. *Lancet* 2004;**364**: 347–354.

- 77. Guazzi M, Vicenzi M, Arena R, Guazzi MD. PDE5 inhibition with sildenafil improves left ventricular diastolic function, cardiac geometry, and clinical status in patients with stable systolic heart failure: results of a 1-year, prospective, randomized, placebo-controlled study. *Circ Heart Fail* 2011;**4**:8–17.
- Guazzi M, Vicenzi M, Arena R, Guazzi MD. Pulmonary hypertension in heart failure with preserved ejection fraction: a target of phosphodiesterase-5 inhibition in a 1-year study. *Circulation* 2011;**124**:164–174.
- 79. Redfield MM, Chen HH, Borlaug BA, Semigran MJ, Lee KL, Lewis G, LeWinter MM, Rouleau JL, Bull DA, Mann DL, Deswal A, Stevenson LW, Givertz MM, Ofili EO, O'Connor CM, Felker GM, Goldsmith SR, Bart BA, McNulty SE, Ibarra JC, Lin G, Oh JK, Patel MR, Kim RJ, Tracy RP, Velazquez EJ, Anstrom KJ, Hernandez AF, Mascette AM, Braunwald E; RELAX Trial. Effect of phosphodiesterase-5 inhibition on exercise capacity and clinical status in heart failure with preserved ejection fraction: a randomized clinical trial. JAWA 2013;309:1268–1277.
- Evgenov OV, Pacher P, Schmidt PM, Hasko G, Schmidt HH, Stasch JP. NO-independent stimulators and activators of soluble guanylate cyclase: discovery and therapeutic potential. *Nat Rev Drug Discov* 2006;5:755–768.
- Stasch JP, Becker EM, Alonso-Alija C, Apeler H, Dembowsky K, Feurer A, Gerzer R, Minuth T, Perzborn E, Pleiss U, Schroder H, Schroeder W, Stahl E, Steinke W, Straub A, Schramm M. NO-independent regulatory site on soluble guanylate cyclase. *Nature* 2001;**410**:212–215.
- Schindler U, Strobel H, Schonafinger K, Linz W, Lohn M, Martorana PA, Rutten H, Schindler PW, Busch AE, Sohn M, Topfer A, Pistorius A, Jannek C, Mulsch A. Biochemistry and pharmacology of novel anthranilic acid derivatives activating heme-oxidized soluble guanylyl cyclase. *Mol Pharmacol* 2006;69:1260–1268.
- Boerrigter G, Costello-Boerrigter LC, Cataliotti A, Lapp H, Stasch JP, Burnett JC Jr. Targeting heme-oxidized soluble guanylate cyclase in experimental heart failure. *Hypertension* 2007;49:1128–1133.
- Kalk P, Godes M, Relle K, Rothkegel C, Hucke A, Stasch JP, Hocher B. NO-independent activation of soluble guanylate cyclase prevents disease progression in rats with 5/6 nephrectomy. *Br J Pharmacol* 2006;**148**:853–859.
- 85. Gheorghiade M, Erdmann E, Ferrari R, Filippatos G, Levy PD, Maggioni A, Mebazaa A, Nowack C. Treatment of acute decompensated heart failure with the soluble guanylate cyclase activator cinaciguat: The COMPOSE program – three randomized, controlled, phase IIb studies. J Card Fail 2011;17:971.
- Masuyama H, Tsuruda T, Sekita Y, Hatakeyama K, Imamura T, Kato J, Asada Y, Stasch JP, Kitamura K. Pressure-independent effects of pharmacological stimulation of soluble guanylate cyclase on fibrosis in pressure-overloaded rat heart. *Hypertens Res* 2009;**32**:597–603.
- Jones E, Kemp-Harper B, Stasch J-P, Schmidt H, Widdop R. Cardioprotective effects in aged spontaneously hypertensive rats due to chronic stimulation/activation of sGC without hypotension. *BMC Pharmacol* 2009;9 (Suppl. 1):P29.
- Sharkovska Y, Kalk P, Lawrenz B, Godes M, Hoffmann LS, Wellkisch K, Geschka S, Relle K, Hocher B, Stasch JP. Nitric oxide-independent stimulation of soluble guanylate cyclase reduces organ damage in experimental low-renin and high-renin models. J Hypertens 2010;28:1666–1675.
- Boerrigter G, Costello-Boerrigter LC, Cataliotti A, Tsuruda T, Harty GJ, Lapp H, Stasch JP, Burnett JC Jr. Cardiorenal and humoral properties of a novel direct soluble guanylate cyclase stimulator BAY 41–2272 in experimental congestive heart failure. *Circulation* 2003;**107**:686–689.
- Mitrovic V, Swidnicki B, Ghofrani A, Muck W, Kirschbaum N, Mittendorf J, Stasch J-P, Wensing G, Frey R, Lentini S. Acute hemodynamic response to single oral doses of BAY 60–4552, a soluble guanylate cyclase stimulator, in patients with biventricular heart failure. *BMC Pharmacol* 2009;9(Suppl. 1):P51.
- 91. Bonderman D, Pretsch I, Steringer-Mascherbauer R, Jansa P, Rosenkranz S, Tufaro C, Bojic A, Lam CS, Frey R, Ochan Kilama M, Unger S, Roessig L, Lang IM. Acute hemodynamic effects of riociguat in patients with pulmonary hypertension associated with diastolic heart failure (DILATE-1): a randomized, double-blind, placebo-controlled, single-dose study. *Chest* 2014;**146**:1274–1285.
- Gu J, Noe A, Chandra P, Al-Fayoumi S, Ligueros-Saylan M, Sarangapani R, Maahs S, Ksander G, Rigel DF, Jeng AY, Lin TH, Zheng W, Dole WP. Pharmacokinetics and pharmacodynamics of LCZ696, a novel dual-acting angiotensin receptor-neprilysin inhibitor (ARNi). J Clin Pharmacol 2010;50:401–414.
- Meulemans AL, Sipido KR, Sys SU, Brutsaert DL. Atriopeptin III induces early relaxation of isolated mammalian papillary muscle. *Circ Res* 1988;62:1171–1174.
- Potter LR, Abbey-Hosch S, Dickey DM. Natriuretic peptides, their receptors, and cyclic guanosine monophosphate-dependent signaling functions. *Endocr Rev* 2006; 27:47–72.
- Gardner DG, Chen S, Glenn DJ, Grigsby CL. Molecular biology of the natriuretic peptide system: implications for physiology and hypertension. *Hypertension* 2007; 49:419–426.
- 96. McMurray JJ, Packer M, Desai AS, Gong J, Lefkowitz MP, Rizkala AR, Rouleau J, Shi VC, Solomon SD, Swedberg K, Zile MR. Dual angiotensin receptor and neprilysin inhibition as an alternative to angiotensin-converting enzyme inhibition in patients with chronic systolic heart failure: rationale for and design of the

Prospective comparison of ARNI with ACEI to Determine Impact on Global Mortality and morbidity in Heart Failure trial (PARADIGM-HF). *Eur J Heart Fail* 2013;**15**: 1062–1073.

- McMurray JJV, Packer M, Desai AS, Gong J, Lefkowitz MP, Rizkala AR, Rouleau JL, Shi VC, Solomon SD, Swedberg K, Zile MR. Angiotensin–neprilysin inhibition versus enalapril in heart failure. N Engl J Med 2014;371:993–1004.
- 98. Solomon SD, Zile M, Pieske B, Voors A, Shah A, Kraigher-Krainer E, Shi V, Bransford T, Takeuchi M, Gong J, Lefkowitz M, Packer M, McMurray JJ, Prospective comparison of ARNI with ARB on Management Of heart failUre with preserved ejectioN fracTion (PARAMOUNT) Investigators. The angiotensin receptor neprilysin inhibitor LCZ696 in heart failure with preserved ejection fraction: a phase 2 double-blind randomised controlled trial. *Lancet* 2012;**380**:1387–1395.
- Bathgate RA, Halls ML, van der Westhuizen ET, Callander GE, Kocan M, Summers RJ. Relaxin family peptides and their receptors. *Physiol Rev* 2013;93: 405–480.
- 100. Jelinic M, Leo CH, Post Uiterweer ED, Sandow SL, Gooi JH, Wlodek ME, Conrad KP, Parkington H, Tare M, Parry LJ. Localization of relaxin receptors in arteries and veins, and region-specific increases in compliance and bradykininmediated relaxation after in vivo serelaxin treatment. FASEB J 2014;28:275–287.
- 101. Teerlink JR, Cotter G, Davison BA, Felker GM, Filippatos G, Greenberg BH, Ponikowski P, Unemori E, Voors AA, Adams KF Jr, Dorobantu MI, Grinfeld LR, Jondeau G, Marmor A, Masip J, Pang PS, Werdan K, Teichman SL, Trapani A, Bush CA, Saini R, Schumacher C, Severin TM, Metra M, RELAXin in Acute Heart Failure (RELAX-AHF) Investigators. Serelaxin, recombinant human relaxin-2, for treatment of acute heart failure (RELAX-AHF): a randomised, placebo-controlled trial. *Lancet* 2013;**381**:29–39.
- 102. Ponikowski P, Metra M, Teerlink JR, Unemori E, Felker GM, Voors AA, Filippatos G, Greenberg B, Teichman SL, Severin T, Mueller-Velten G, Cotter G, Davison BA. Design of the RELAXin in acute heart failure study. Am Heart J 2012; 163:149–55 e1.
- Conrad KP. Unveiling the vasodilatory actions and mechanisms of relaxin. Hypertension 2010;56:2–9.
- 104. Dschietzig T, Brecht A, Bartsch C, Baumann G, Stangl K, Alexiou K. Relaxin improves TNF-alpha-induced endothelial dysfunction: the role of glucocorticoid receptor and phosphatidylinositol 3-kinase signalling. *Cardiovasc Res* 2012;**95**: 97–107.
- Leo CH, Jelinic M, Parkington HC, Tare M, Parry LJ. Acute intravenous injection of serelaxin (recombinant human relaxin-2) causes rapid and sustained bradykininmediated vasorelaxation. J Am Heart Assoc 2014;3:e000493.
- McGuane JT, Debrah JE, Sautina L, Jarajapu YP, Novak J, Rubin JP, Grant MB, Segal M, Conrad KP. Relaxin induces rapid dilation of rodent small renal and human subcutaneous arteries via PI3 kinase and nitric oxide. *Endocrinology* 2011;**152**:2786–2796.
- Segal MS, Sautina L, Li S, Diao Y, Agoulnik AI, Kielczewski J, McGuane JT, Grant MB, Conrad KP. Relaxin increases human endothelial progenitor cell NO and migration and vasculogenesis in mice. *Blood* 2012;**119**:629–636.
- Diez J. Serelaxin: a novel therapy for acute heart failure with a range of hemodynamic and non-hemodynamic actions. Am J Cardiovasc Drugs 2014;14:275–285.
- Hornig B, Maier V, Drexler H. Physical training improves endothelial function in patients with chronic heart failure. *Circulation* 1996;93:210–214.
- Kojda G, Hambrecht R. Molecular mechanisms of vascular adaptations to exercise. Physical activity as an effective antioxidant therapy? *Cardiovasc Res* 2005;67: 187–197.
- 111. Mo M, Eskin SG, Schilling WP. Flow-induced changes in Ca²⁺ signaling of vascular endothelial cells: effect of shear stress and ATP. Am J Physiol 1991;**260**(Pt 2): H1698–H1707.
- Olesen SP, Clapham DE, Davies PF. Haemodynamic shear stress activates a K⁺ current in vascular endothelial cells. *Nature* 1988;**331**:168–170.
- 113. O'Connor CM, Whellan DJ, Lee KL, Keteyian SJ, Cooper LS, Ellis SJ, Leifer ES, Kraus WE, Kitzman DW, Blumenthal JA, Rendall DS, Miller NH, Fleg JL, Schulman KA, McKelvie RS, Zannad F, Pina IL. Efficacy and safety of exercise training in patients with chronic heart failure: HF-ACTION randomized controlled trial. JAMA 2009;301:1439–1450.
- 114. Hamburg NM, Palmisano J, Larson MG, Sullivan LM, Lehman BT, Vasan RS, Levy D, Mitchell GF, Vita JA, Benjamin EJ. Relation of brachial and digital measures of vascular function in the community: the Framingham heart study. *Hypertension* 2011;**57**: 390–396.
- 115. Piña IL, Bittner V, Clare RM, Swank A, Kao A, Safford R, Nigam A, Barnard D, Walsh MN, Ellis SJ, Keteyian SJ. Effects of exercise training on outcomes in women with heart failureanalysis of HF-ACTION (Heart Failure–A Controlled Trial Investigating Outcomes of Exercise TraiNing) by Sex. JACC Heart Fail 2014; 2:180–186.
- 116. Edelmann F, Gelbrich G, Dungen HD, Frohling S, Wachter R, Stahrenberg R, Binder L, Topper A, Lashki DJ, Schwarz S, Herrmann-Lingen C, Loffler M, Hasenfuss G, Halle M, Pieske B. Exercise training improves exercise capacity and diastolic function in patients with heart failure with preserved ejection fraction:

results of the Ex-DHF (Exercise training in Diastolic Heart Failure) pilot study. J Am Coll Cardiol 2011;**58**:1780–1791.

- 117. Kitzman DW, Brubaker PH, Morgan TM, Stewart KP, Little WC. Exercise training in older patients with heart failure and preserved ejection fraction: a randomized, controlled, single-blind trial. *Circ Heart Fail* 2010;**3**:659–667.
- 118. Kitzman DW, Brubaker PH, Herrington DM, Morgan TM, Stewart KP, Hundley WG, Abdelhamed A, Haykowsky MJ. Effect of endurance exercise training on endothelial function and arterial stiffness in older patients with heart failure and preserved ejection fraction: a randomized, controlled, single-blind trial. J Am Coll Cardiol 2013;62:584–592.
- 119. Gao R, Zhang J, Cheng L, Wu X, Dong W, Yang X, Li T, Liu X, Xu Y, Li X, Zhou M. A Phase II, randomized, double-blind, multicenter, based on standard therapy, placebo-controlled study of the efficacy and safety of recombinant human neuregulin-1 in patients with chronic heart failure. J Am Coll Cardiol 2010;55: 1907–1914.
- 120. Jabbour A, Hayward CS, Keogh AM, Kotlyar E, McCrohon JA, England JF, Amor R, Liu X, Li XY, Zhou MD, Graham RM, Macdonald PS. Parenteral administration of recombinant human neuregulin-1 to patients with stable chronic heart failure produces favourable acute and chronic haemodynamic responses. *Eur J Heart Fail* 2011; **13**:83–92.
- 121. Jay SM, Murthy AC, Hawkins JF, Wortzel JR, Steinhauser ML, Alvarez LM, Gannon J, Macrae CA, Griffith LG, Lee RT. An engineered bivalent neuregulin protects against doxorubicin-induced cardiotoxicity with reduced proneoplastic potential. *Circulation* 2013;**128**:152–161.
- 122. Amabile N, Cheng S, Renard JM, Larson MG, Ghorbani A, McCabe E, Griffin G, Guerin C, Ho JE, Shaw SY, Cohen KS, Vasan RS, Tedgui A, Boulanger CM, Wang TJ. Association of circulating endothelial microparticles with cardiometabolic risk factors in the Framingham Heart Study. *Eur Heart J* 2014;**35**:2972–2979.
- Marti CN, Gheorghiade M, Kalogeropoulos AP, Georgiopoulou VV, Quyyumi AA, Butler J. Endothelial dysfunction, arterial stiffness, and heart failure. J Am Coll Cardiol 2012;60:1455–1469.
- Shantsila E, Wrigley BJ, Blann AD, Gill PS, Lip GY. A contemporary view on endothelial function in heart failure. *Eur J Heart Fail* 2012;**14**:873–881.
- 125. Galindo CL, Kasasbeh E, Murphy A, Ryzhov S, Lenihan S, Ahmad FA, Williams P, Nunnally A, Adcock J, Song Y, Harrell FE, Tran TL, Parry TJ, Iaci J, Ganguly A, Feoktistov I, Stephenson MK, Caggiano AO, Sawyer DB, Cleator JH. Antiremodeling and anti-fibrotic effects of the neuregulin-1beta glial growth factor 2 in a large animal model of heart failure. J Am Heart Assoc 2014;**3**:e000773.
- 126. von Lueder TG, Wang BH, Kompa AR, Huang L, Webb R, Jordaan P, Atar D, Krum H. The Angiotensin-Receptor Neprilysin Inhibitor LCZ696 Attenuates Cardiac Remodeling and Dysfunction After Myocardial Infarction by Reducing Cardiac Fibrosis and Hypertrophy. *Circ Heart Fail* 2015;8:71–78.
- Brunner F, Brás-Silva C, Cerdeira AS, Leite-Moreira AF. Cardiovascular endothelins: essential regulators of cardiovascular homeostasis. *Pharmacol Ther* 2006;**111**: 508–531.
- 128. Hara A, Yuhki K, Fujino T, Yamada T, Takayama K, Kuriyama S, Takahata O, Karibe H, Okada Y, Xiao CY, Ma H, Narumiya S, Ushikubi F. Augmented cardiac hypertrophy in response to pressure overload in mice lacking the prostaglandin I2 receptor. *Circulation* 2005;**112**:84–92.
- Ritchie RH, Rosenkranz AC, Huynh LP, Stephenson T, Kaye DM, Dusting GJ. Activation of IP prostanoid receptors prevents cardiomyocyte hypertrophy via cAMP-dependent signaling. *Am J Physiol Heart Circ Physiol* 2004;287:H1179–H1185.
- Lemmens K, Fransen P, Sys SU, Brutsaert DL, De Keulenaer GW. Neuregulin-1 induces a negative inotropic effect in cardiac muscle: role of nitric oxide synthase. *Circulation* 2004;**109**:324–326.
- Lemmens K, Doggen K, De Keulenaer GW. Activation of the neuregulin/ErbB system during physiological ventricular remodeling in pregnancy. Am J Physiol Heart Circ Physiol 2011;300:H931–H942.
- 132. Ladage D, Yaniz-Galende E, Rapti K, Ishikawa K, Tilemann L, Shapiro S, Takewa Y, Muller-Ehmsen J, Schwarz M, Garcia MJ, Sanz J, Hajjar RJ, Kawase Y. Stimulating myocardial regeneration with periostin peptide in large mammals improves function post-myocardial infarction but increases myocardial fibrosis. *PLoS One* 2013; 8:e59656.
- Kuhn B, del Monte F, Hajjar RJ, Chang YS, Lebeche D, Arab S, Keating MT. Periostin induces proliferation of differentiated cardiomyocytes and promotes cardiac repair. Nat Med 2007;13:962–969.
- 134. Cingolani OH, Kirk JA, Seo K, Koitabashi N, Lee D-i, Ramirez-Correa G, Bedja D, Barth AS, Moens AL, Kass DA. Thrombospondin-4 is required for stretchmediated contractility augmentation in cardiac muscle. *Circ Res* 2011;**109**: 1410–1414.
- 135. Frolova EG, Sopko N, Blech L, Popović ZB, Li J, Vasanji A, Drumm C, Krukovets I, Jain MK, Penn MS, Plow EF, Stenina OI. Thrombospondin-4 regulates fibrosis and remodeling of the myocardium in response to pressure overload. *FASEB J* 2012; 26:2363–2373.

- 136. Swinnen M, Vanhoutte D, Van Almen GC, Hamdani N, Schellings MWM, D'hooge J, Van der Velden J, Weaver MS, Sage EH, Bornstein P, Verheyen FK, VandenDriessche T, Chuah MK, Westermann D, Paulus WJ, Van de Werf F, Schroen B, Carmeliet P, Pinto YM, Heymans S. Absence of thrombospondin-2 causes age-related dilated cardiomyopathy. *Circulation* 2009;**120**:1585–1597.
- 137. Xia Y, Dobaczewski M, Gonzalez-Quesada C, Chen W, Biernacka A, Li N, Lee D-W, Frangogiannis NG. Endogenous thrombospondin 1 protects the pressure-overloaded myocardium by modulating fibroblast phenotype and matrix metabolism. *Hypertension* 2011;**58**:902–911.
- Oshima Y, Ouchi N, Sato K, Izumiya Y, Pimentel DR, Walsh K. Follistatin-like 1 is an Akt-regulated cardioprotective factor that is secreted by the heart. *Circulation* 2008; 117:3099–3108.
- Shimano M, Ouchi N, Nakamura K, Oshima Y, Higuchi A, Pimentel DR, Panse KD, Lara-Pezzi E, Lee S-J, Sam F, Walsh K. Cardiac myocyte-specific ablation of follistatin-like 3 attenuates stress-induced myocardial hypertrophy. J Biol Chem 2011;286:9840–9848.
- 140. Shimano M, Ouchi N, Nakamura K, van Wijk B, Ohashi K, Asaumi Y, Higuchi A, Pimentel DR, Sam F, Murohara T, van den Hoff MJB, Walsh K. Cardiac myocyte follistatin-like 1 functions to attenuate hypertrophy following pressure overload. *Proc Natl Acad Sci USA* 2011;**108**:E899–E906.
- 141. Oshima Y, Ouchi N, Shimano M, Pimentel DR, Papanicolaou KN, Panse KD, Tsuchida K, Lara-Pezzi E, Lee S-J, Walsh K. Activin A and follistatin-like 3 determine the susceptibility of heart to ischemic injury. *Circulation* 2009;**120**: 1606–1615.

- 142. Hayata N, Fujio Y, Yamamoto Y, Iwakura T, Obana M, Takai M, Mohri T, Nonen S, Maeda M, Azuma J. Connective tissue growth factor induces cardiac hypertrophy through Akt signaling. *Biochem Biophys Res Commun* 2008;**370**:274–278.
- 143. Panek AN, Posch MG, Alenina N, Ghadge SK, Erdmann B, Popova E, Perrot A, Geier C, Morano RDI, Bader M, Özcelik C. Connective tissue growth factor overexpression in cardiomyocytes promotes cardiac hypertrophy and protection against pressure overload. *PLoS One* 2009;**4**:e6743.
- 144. Zhang Y, Liu Y, Zhu XH, Zhang XD, Jiang DS, Bian ZY, Zhang XF, Chen K, Wei X, Gao L, Zhu LH, Yang Q, Fan GC, Lau WB, Ma X, Li H. Dickkopf-3 attenuates pressure overload-induced cardiac remodelling. *Cardiovasc Res* 2014;**102**:35–45.
- 145. Perjés Á, Skoumal R, Tenhunen O, Kónyi A, Simon M, Horváth IG, Kerkelä R, Ruskoaho H, Szokodi I. Apelin increases cardiac contractility via protein kinase Cε- and extracellular signal-regulated kinase-dependent mechanisms. *PLoS One* 2014;**9**:e93473.
- Ceylan-Isik AF, Kandadi MR, Xu X, Hua Y, Chicco AJ, Ren J, Nair S. Apelin administration ameliorates high fat diet-induced cardiac hypertrophy and contractile dysfunction. J Mol Cell Cardiol 2013;63:4–13.
- 147. Ashley EA, Powers J, Chen M, Kundu R, Finsterbach T, Caffarelli A, Deng A, Eichhorn J, Mahajan R, Agrawal R, Greve J, Robbins R, Patterson AJ, Bernstein D, Quertermous T. The endogenous peptide apelin potently improves cardiac contractility and reduces cardiac loading in vivo. *Cardiovasc Res* 2005;65:73–82.
- 148. Halkein J, Tabruyn SP, Ricke-Hoch M, Haghikia A, Nguyen NQ, Scherr M, Castermans K, Malvaux L, Lambert V, Thiry M, Sliwa K, Noel A, Martial JA, Hilfiker-Kleiner D, Struman I. MicroRNA-146a is a therapeutic target and biomarker for peripartum cardiomyopathy. *J Clin Invest* 2013;**123**:2143–2154.